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## Chitosan and *Opuntia ficus-indica* mucilage as the base of a polymeric edible film for the protection of tomatoes against *Rhizopus stolonifer*

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### ABSTRACT

Chitosan (CS) is a versatile raw material for the production of fibers and biomaterials. As antifungal compound, it has an interesting mechanism related to interactions with the cell membranes, being effective against various fungi. Likewise, *Opuntia* mucilage (OM) is a polymeric matrix with interesting applications in the agricultural and food field. An edible film based on these materials was designed in this work, as an alternative to defend crops against fungi infections. Various tests of homogeneity and resistance were performed by varying the concentration of chitosan, glycerol and mucilage to select the best composition for the film. Subsequently, structural and rheological studies were carried out as elements for the characterization of the chosen formulation. Finally, *in vitro* and *in situ* tests were performed to evaluate the antifungal potential in the protecting the crops of interest. Based on the qualitative and physicochemical characterization, it was determined that the film with better integrity in its structure was obtained at concentrations of 3% glycerol, 2% chitosan and 20% mucilage. The resulting film was homogenous, flexible, luminous, slightly dark, with cumulative viscosity, that means that total viscosity is the result of each component viscosity. The CS-OM film demonstrated a strong antifungal effect against *Rhizopus stolonifer* *in vitro* and *in situ* conditions and increased the shelf-life of tomatoes. The results showed the potential of the use of this biodegradable film in protecting the crops of interest, making it an attractive polymeric product for field application.

**Key Words:** Chitosan, *Opuntia* mucilage, edible film, *Rhizopus stolonifer*.

### Quitosano y mucílago de *Opuntia ficus-indica* (nopal) como base de una película polimérica comestible para la protección de tomates contra *Rhizopus stolonifer*

### RESUMEN

El quitosano es una materia prima extremadamente versátil para la producción de fibras y biomateriales. Como antifúngico, posee un mecanismo de acción relacionado con interacciones electrostáticas de las membranas celulares, siendo efectivo contra diversos hongos. Por otro lado, el mucílago de nopal (*Opuntia*) es una matriz polimérica con aplicaciones interesantes en el campo agrícola y alimentario como componente estructural de geles o suplemento dietético. En este trabajo se diseñó una película comestible basada en quitosano (antifúngico) y mucílago de nopal (estructurante) como una alternativa para la defensa de los cultivos contra las infecciones por hongos. Se realizaron diversas pruebas de homogeneidad y resistencia variando la concentración de quitosano, glicerol y mucílago para seleccionar la mejor composición para la película. Posteriormente se realizaron estudios estructurales y reológicos como elementos para la caracterización de la formulación elegida. Finalmente, se hicieron pruebas *in vitro* e *in situ* para evaluar el potencial antifúngico en la protección de frutos de interés. Con base en la caracterización cualitativa y fisicoquímica se determinó que la película con integridad homogénea en su estructura se obtuvo a concentraciones de 3% de glicerol, 2% de quitosano y 20% de mucílago de nopal. La película resultante tuvo una superficie homogénea, flexible, luminosa, ligeramente oscura y con viscosidad acumulativa, lo que significa que la viscosidad total es la suma de las viscosidades de cada componente. La película demostró un fuerte efecto antifúngico contra *Rhizopus stolonifer* en condiciones *in vitro* e *in situ* y aumentó la vida útil de los frutos probados. Los resultados mostraron el potencial del uso de esta película ecológica para la protección de frutas de interés, lo que la convierte en un producto polimérico atractivo para su aplicación en el campo.

**Palabras Clave:** Quitosano, mucílago de nopal, película comestible, *Rhizopus stolonifer*.

## INTRODUCTION

**A**s a result of the increase in the human population, agriculture techniques must be improved to face the challenge of feeding humanity with healthy and nutritious food, exchanging ancient methods for the use of chemical substances that enhance growth and reduce losses due to pests (Hahn, 2014). Moreover, the trend of eating fresh products for an appropriated diet has consequently led to food being processed faster, which increases the risk of contamination by pesticides, toxins, or microorganisms (Jung, Jang & Matthews, 2014).

In the context of agricultural production, there are risks associated with the loss of crops at all stages of the process. However, post-harvest losses are one of the most significant since they occur immediately prior to the product reaching the consumer (Kasso & Bekele, 2018). Post-harvest activities represent 34% of global crop losses, with the most frequent causes being pests, natural ripening processes, environmental conditions and microbial infection mainly with fungi (Abass *et al.*, 2014; Kasso & Bekele, 2018; Kumar & Kalita, 2017).

The infection of harvested crops with phytopathogenic fungi represents one of the most important challenges in current agricultural techniques, not only from an economic point of view but for the risk to human health. Several factors are related to fungal infection under post-harvest conditions, such as pH, temperature, UV and oxidative stress, among others (Liu *et al.*, 2018). In recent decades, the use of organic fungicides (such as site-specific fungicides, thiabendazole, pyrimethanil, o-phenylphenol and imazalil), has been the main force in avoiding crop decay due to their effectiveness and spectrum of action. However, the use of these substances has a serious environmental impact and causes human health disorders and antimicrobial resistance (Hahn, 2014; Peris-Vicente, Marzo-Mas, Roca-Genovés, Carda-Broch & Esteve-Romero, 2016).

The current trends in crop protection involve a range of actions that mitigate the environmental impact, have a broad spectrum of action and are of natural origin, biodegradable and easy to apply (Seiber, Coats, Duke & Gross, 2014). One alternative is the use of natural materials such as chitosan and *Opuntia mucilage* as edible film presentation.

Chitosan (CS) is one of the most promising resources for the generation of bioactive, biodegradable materials against infections that affect crops of economic importance. The main chain of this semi-natural polymer displays a polycationic behavior in acidic solution due to the free amino groups in its structure (Olicón-Hernández, Uribe-Álvarez, Uribe-Carvajal, Pardo & Guerra-Sánchez, 2017). This polymer is obtained by the deacetylation of chitin, the second most abundant structural

polymer related to fungi, crustaceans and insects (Olicón-Hernández, Zepeda Giraud & Guerra-Sánchez, 2017). Due to its properties, such as resistance, flexibility, polymerization, and polycationic charge, chitosan has been used as a raw material for the formation of gels, fibers and films and has been shown to have extensive antimicrobial activity against bacteria and fungi (Anitha *et al.*, 2014; Olicón-Hernández *et al.*, 2015; Olicón-Hernández, Zepeda Giraud, *et al.*, 2017; Younes & Rinaudo, 2015).

In the context of the edible films formulation, a typical edible film has three major components; film forming material, plasticizer, and additives. From this point of view Nopal (*Opuntia ficus-indica*) is a potential alternative as element of edible film. The nopal is a traditional element of Mexican food and culture but is considered an exotic ingredient in an international context (de Albuquerque *et al.*, 2018). Extracts of this plant have shown benefits to human health, such as hypolipidemic, hypocholesterolemic, antidiabetic, hypoglycemic, antioxidant and anti-inflammatory efficacies (Otálora, Carriazo, Iturriaga, Nazareno & Osorio, 2015). *Opuntia mucilage* (OM) is a complex material formed from carbohydrates including L-arabinose, D-galactose, D-xylose, L-rhamnose and D-galacturonic acid in variable proportions, which has been cataloged as a promising natural resource for use in the food, pharmaceutical and construction industries (León-Martínez, Cano-Barrita, Lagunez-Rivera & Medina-Torres, 2014; Otálora *et al.*, 2015).

In the present study, we designed a CS-OM film for the protection of crops, which has the characteristic of being edible, allowing safe and practical handling for its application. Physicochemical characterization of the film was performed and the *in vitro* and *in situ* antifungal activity against *Rhizopus stolonifer*, one of the most important phytopathogenic fungi, was evaluated.

## MATERIALS AND METHODS

### Reagents and Solutions

Low-molecular weight chitosan (50,000–190,000 Da) was provided by Sigma–Aldrich (St. Louis, MO, USA). A stock solution of 10 mg/mL was made in 1% acetic acid, according to our previously reported protocol, followed by sterilization via autoclaving (Olicón-Hernández *et al.*, 2015).

The *Opuntia* (nopal) cladodes for OM extraction were obtained from a local market in México City. The cladodes were thoroughly washed, cut into uniform pieces (0.5 cm x 0.5 cm), placed in water (5:1 w/v) and incubated overnight at room temperature. Subsequently, the mixture was filtered and concentrated by evaporation.

Glycerol and gelatin were of commercial food grade and purchased from a local supermarket.

### CS-OM film formulation

The film formulation was made by mixing four main elements: 1) CS, 2) OM, 3) glycerol and 4) gelatin. The proportions of OM and gelatin kept constant (20 and 10%, respectively) to maintain the integrity of the membrane. Eight different preparations were made, with two concentrations of glycerol (1 and 3%), to evaluate the flexibility and four concentrations of chitosan (0.05, 0.1, 0.15 and 0.20%), to test the homogeneity and antifungal effect of the resulting film (Table I). Each mixture was homogenized using a vortex and placed into Petri dishes (8.7 cm), at room temperature until dry. The film was manually separated from the plate. Selection was made based on the integrity, flexibility and handling of the resulting film, in addition to the homogeneity, as confirmed by conventional microscopy.

### Scanning electron microscopy (SEM)

The selected film was cut into 1cm x 1cm pieces and placed on metallic bases for structural evaluation by SEM. After the samples were dried, a gold layer was directly applied under a vacuum for 10 min and observed using a JEOL 5800LV scanning electron microscope (Tokyo, Japan) at 15 kV (Olicón-Hernández *et al.*, 2015).

### Physicochemical characterization

#### Viscosity

The viscosity of the selected formulation was determined as a characterization element. Likewise, the viscosity of each of the elements of the mixture was determined separately to evaluate its rheological behavior. A viscometer Brookfield model RVT Spring Torque (Dyne-cm), 7,187.0 (Middleboro, MA, USA), was used. Six spindles and eight velocities were tested to standardize the determination of each component (manual instructions). A volume of 600 mL sample was placed in a beaker and the spindle of the viscometer was submerged and centered in the sample. Rotation of the viscometer was maintained until the reading stabilized and the determination of viscosity was carried out according to the specifications described in the manual (Viscometer, 2016). The experiments were performed in triplicate and the viscosity is expressed in centipoises (cp).

#### Color

The color determination of the film was carried out using a CR-10 colorimeter (Konica Minolta, Japan). The determination was randomly performed on the surface of three different samples and the L\*, a\* and b\* values were measured based on the CIE Lab\* color space (Valadez-Carmona *et al.*, 2016).

#### Percentage humidity and film solubility in water

The humidity was measured by gravimetric standard methods and is expressed as a percentage (%). The experiments were performed in triplicate. Percentage solubility in water was measured according to García *et al.*, 2004, based on the

comparison of the weight of the dry sample prior to and following exhaustive agitation in water, as calculated using the following equation (García, Pinotti, Martino & Zaritzky, 2004):

$$\% \text{ Solubility} = \left( \frac{\text{Initial dry weight} - \text{Final dry weight}}{\text{Initial dry weight}} \right) * 100$$

### Qualitative evaluation of the protection of stored fruits

Ten Saladette tomatoes (*Lycopersicon esculentum*) were selected according to color, state of health, size and firmness. These fruits were divided into two equal groups: one as a control without protection and the other protected by adhesion of the CS-OM on the surface. The two groups were stored at room temperature and humidity for 30 days. No phytopathogenic agent was intentionally added. After this time, a visual evaluation was made to determine the protective effect of the film as compared with the control.

### Antifungal activity

*Rhizopus stolonifer* R3 was isolated from post-harvest contaminated crops and also provided by CEPROBI-IPN, Yautepec, Morelos, Mexico (Hernández-Lauzardo *et al.*, 2008).

### Determination of *in vitro* antifungal activity

*Rhizopus stolonifer* R3 was grown on commercial potato dextrose agar (PDA) plates for 72 h. After this time, approximately 1-cm<sup>2</sup> pieces of mycelia-agar were excised to inoculate three PDA plates without CS-OM film (control system, Db) and three plates coated with the film (positive system, Da). The plates were incubated at 28 °C until the fungus in the control system covered the plate. In both systems, the diameter of the mycelia was measured to determine the antifungal index according to the following equation (Guo *et al.*, 2006):

$$\text{Antifungal Index (\%)} = \left( 1 - \frac{Da}{Db} \right) * 100$$

where Da is the diameter of the mycelia in the positive system and Db is the diameter in the control system.

### Determination of *in situ* antifungal activity

*R. stolonifer* R3 was grown on PDA plates for 72 h at 28 °C. After this time, a spore suspension was made by mechanical extraction in water. The number of spores was calculated using the Neubauer chamber method and adjusted to 1x10<sup>6</sup> spores/mL in sterile water (Olicón-Hernández, Camacho-Morales, Pozo, González-López & Aranda, 2019).

A total of 90 Saladette tomatoes (*Lycopersicon esculentum*) were selected using the same parameters as described in section 2.5. These tomatoes were extensively washed, disinfected

with 1% sodium hypochlorite and divided into six groups of 15 samples. Three of these groups were protected with the CS-OM film (test groups) and three groups were the control. The spore solution was sprinkled on all tomatoes, which were subsequently placed in humid chambers at room temperature for 72 h. After this time, the severity index (damage) was calculated according to the protocol-scale-equation described by Mayek-Pérez *et al.*, 1995, based on the ratio of samples with visible symptoms of infection to the total samples in the group (Mayek-Pérez, Pedroza-Flores, Villarreal-García & Valdés-Lozano, 1995).

## RESULTS

### Selection of the CS-OM film

Evaluation of the different formulations is displayed in Table I. In general, a low proportion of glycerol resulted in lumpy films with a lack of continuity, which did not allow complete drying. On the other hand, micrographs of the films (Figure 1) revealed that by increasing the concentration of chitosan, the surface was more homogeneous and the flexibility was improved. According to these results, the formulation containing 3% glycerol and 2% chitosan was selected, with the proportions of gelatin and mucilage as specified in section 2.2.

Formulation	Chitosan (%)	Glycerol (%)	Integrity	Flexibility	Homogeneity
1	0.05	1.0	+	+	+
2	0.10		+	++	+
3	0.15		++	++	+
4	0.20		+++	+++	+
5	0.05	3.0	++	++	++
6	0.10		++	++	+++
7	0.15		+++	++	++++
8	0.20		++++	++++	++++

+ = Qualitative evaluation scale. A greater number of symbols represents an increase in the characteristic evaluated.

Table I. Evaluation of the physical characteristics of the different formulations of CS-OM film.

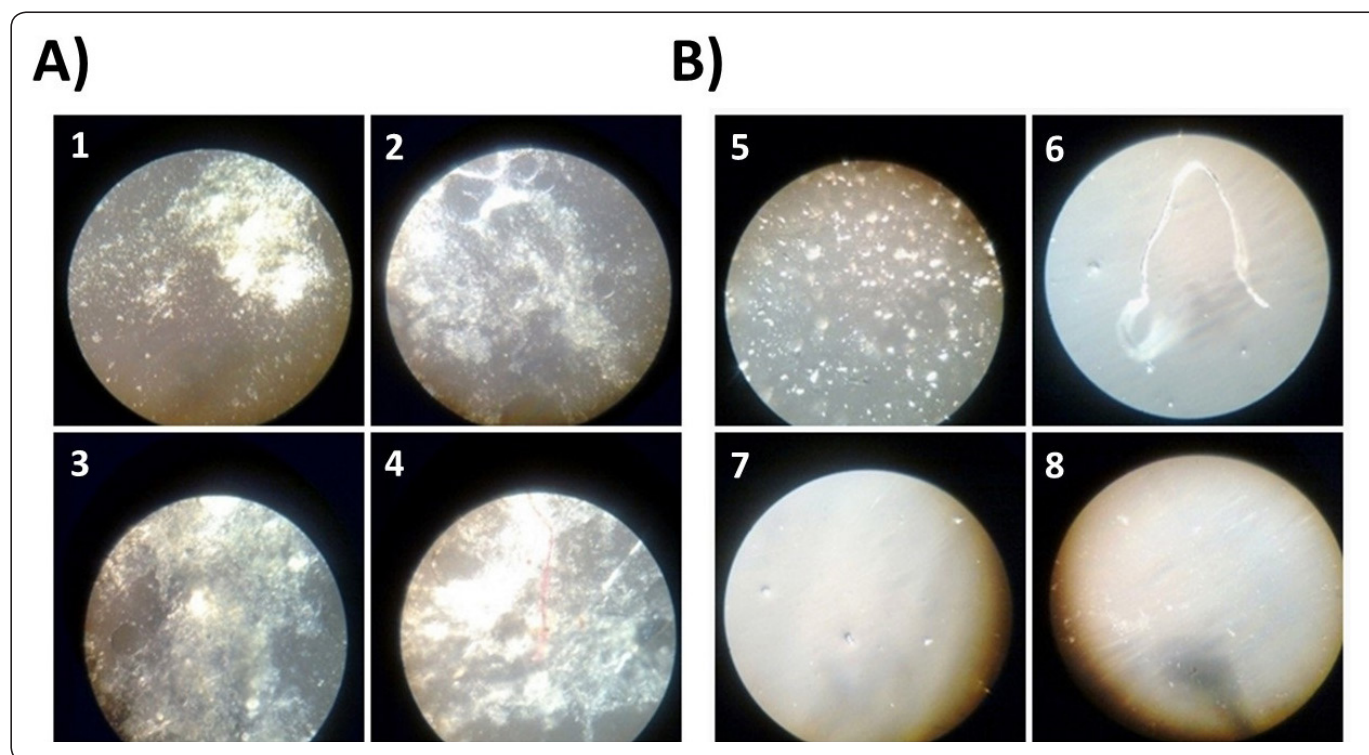


Figure 1. Micrographs of the CS-OM film formulations (100x). Evaluation of the properties of the film was carried out qualitatively. (A) 1% glycerol; (B) 3% glycerol. 1 and 5 = 0.5% chitosan; 2 and 6 = 1% chitosan; 3 and 7 = 1.5% chitosan; 4 and 8 = 2% chitosan.

### Characterization of the CS-OM film

According to SEM analysis, the CS-OM film had a fibrous structure with the absence of pores and a homogenous appearance throughout its surface (Figure 2), which is in accordance with the simple light microscopy data.

The data obtained from the individual elements and the aqueous phase of the CS-OM film (Figure 3), demonstrated that the viscosity was cumulative, with glycerol as the most important element affecting this property.

The data corresponding to humidity, solubility and color are summarized in Table II. Following drying, the resulting film had an average of 12.5% and 43% humidity and solubility, respectively. According to the scales of the L\*, a\* and b\* parameters, the CS-OM film was slightly dark with a light green-blue coloration.

### Effect on the protection of stored fruits

Changes in tomatoes with and without (control) CS-OM film after 30 days of storage are shown in Figure 4. In the control, it was observed that the fruits without protection lost firmness and had slight changes in color, in addition to there being clear evidence of fungal disease. It should be noted that for this determination, no phytopathogenic agent was intentionally added. In contrast, the protected fruits maintained uniform coloration and firmness comparable to the tomatoes under initial storage conditions and no symptoms of the microbial disease were observed.

### *In vitro* and *in situ* antifungal activity

Changes in the growth of *R. stolonifer* with and without CS-OM film on the PDA plates and determination of the antifungal index (%) are shown in Figure 5. The presence of the CS-OM film inhibited the growth of the phytopathogenic agent by approximately 50%. This difference was statistically validated using Tukey's test ( $P < 0.001$ ).

In the case of *in situ* antifungal activity, the tomatoes in the control group (Figure 6A), showed an abundant growth of fungus, which in some cases covered more than 50% of the total surface. During this stage of infection, symptoms of the disease or important changes in coloration or form were not appreciated. However, the severity index in this group reached the maximum value on the scale, indicating that more than 75% of the total fruits were affected.

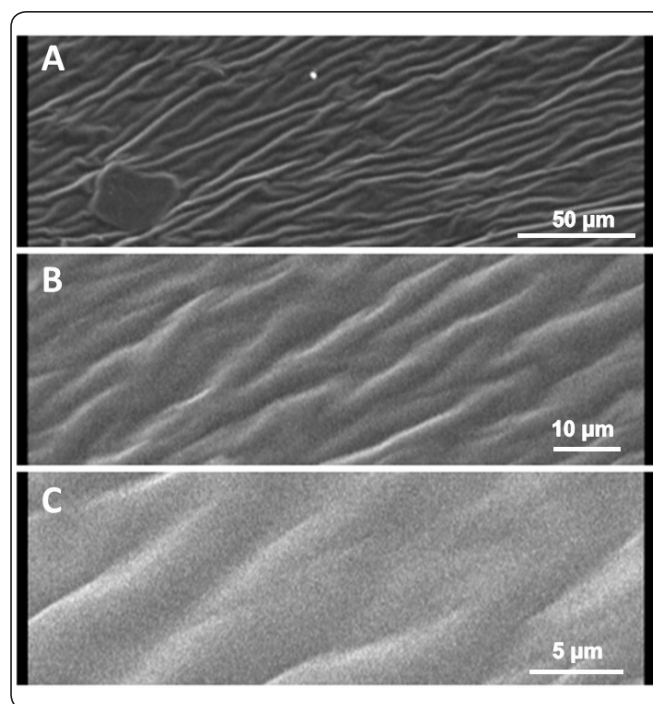


Figure 2. SEM analysis of the CS-OM film. No damage or irregularity was observed in the structure of the film following the drying process. The fibrous structure originated from the interaction of the different components, with chitosan conferring that property. (A) 500x; (B) 1500x; (C) 4000x.

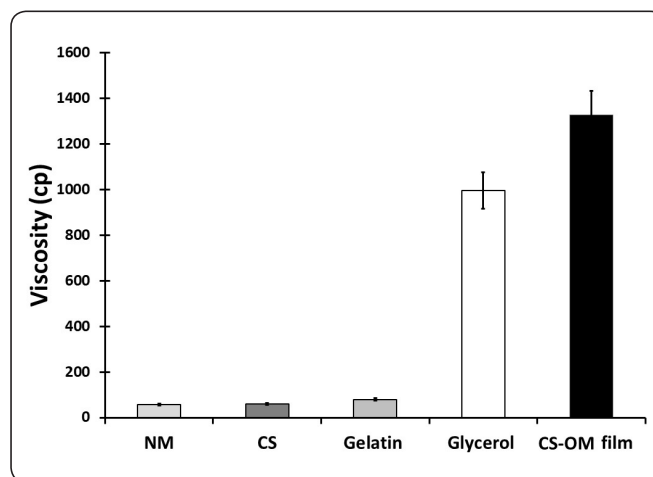


Figure 3. Viscosity determination of the CS-OM film elements. The viscosity is expressed in centipoises. OM = *Opuntia mucilage*; CS = Chitosan; CS-OM film = aqueous phase of the designed film.

Humidity (%)	Solubility (%)	Color					
		White calibration			Black calibration		
		L*	a*	b*	L*	a*	b*
12.4 ± 0.1	43.6 ± 0.1	43.8 ± 3.81	-3.1 ± 0.42	9.0 ± 1.97	32.9 ± 0.84	-1.45 ± 0.35	4.05 ± 0.07

Table II. Percentages of humidity and solubility and color parameters of CS-OM films.

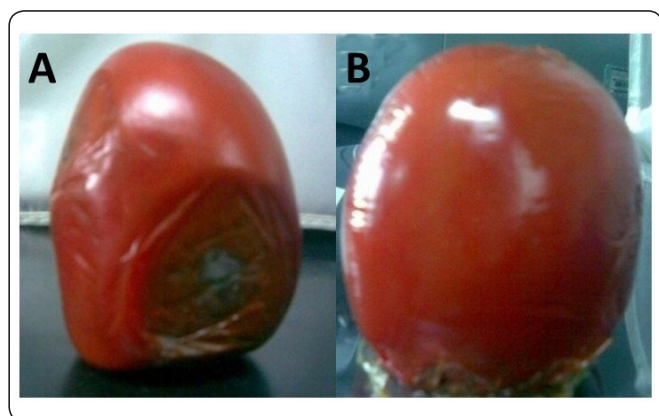


Figure 4. Changes in stored fruits with and without CS-OM film. (A, Control) Fruits without CS-OM film (B) and fruits protected by CS-OM film. The fruits were stored for 30 days. In control fruits, changes in color, texture and firmness were observed and fungal mycelia were present on the surface.

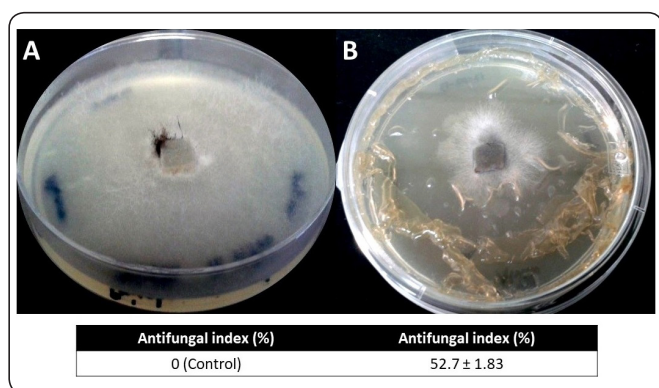


Figure 5. *In vitro* antifungal effect of CS-OM film against *R. stolonifer*. Mycelial growth without film (A, Control) and with the protection of film (B). The fungus took 36 hours to invade the control plates. There was a statistically significant difference validated by Tukey's test ( $P < 0.001$ ).

On the other hand, the fruits protected by the CS-OM film (Figure 6B), obtained a severity index of 2, indicating that less than 25% of the fruits were affected to some degree. Growth in the area of affected fruits was lower and limited to specific areas.

## DISCUSSION

The use of edible films for the protection of crops, fruits and/or meat is a current trend with a view to increasing shelf-life, maximizing productivity and maintaining the quality and flavors of food (Zambrano-Zaragoza *et al.*, 2018). In general, the composition of edible biofilms requires a protein component, a plasticizing agent and in some cases a structural component and/or bioactive agent (Galus & Kadzińska, 2015). In the present work, gelatin and glycerol were selected as the protein and plasticizing agent, respectively, since it has been reported that the combination of these components improves

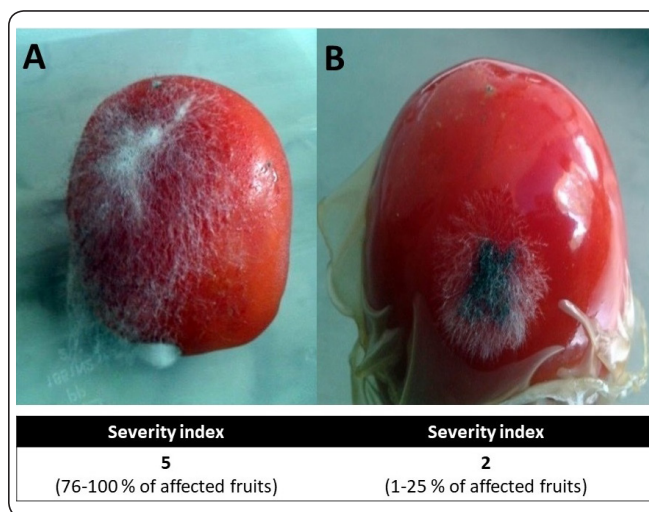


Figure 6. *In situ* antifungal effect of CS-OM film against *R. stolonifer*. Fruits without film (A, Control), had massive mycelial growth on the surface. On the tomatoes protected by CS-OM film (B), a lower and limited fungal growth was observed.

the quality of film and increases the shelf-life of fruits such as avocado and strawberries (Aguilar-Méndez, Martín-Martínez, Tomás, Cruz-Orea, & Jaime-Fonseca, 2008; Del-Valle, Hernández-Muñoz, Guarda & Galotto, 2005).

Several components have been used as elements in film formulation to improve the flexibility and integrity of the coating. Casein, collagen, lipids, soy, corn, alginate, cellulose and essential oils are substances that have been proven to be efficient structural, gelling and active agents (Galus & Kadzińska, 2015). In the present work, the use of *Opuntia* mucilage, an unconventional cheap raw material that contributes carbohydrates, as well as structural and functional elements that make the film edible and complement its biological activity, were explored. In this context, the use of *Opuntia* mucilage as an element in edible films increases water vapor permeability (Domínguez-Martínez *et al.*, 2017), maintains the quality of fruits and vegetables (Allegra *et al.*, 2016; Del-Valle *et al.*, 2005; Treviño-Garza, García, Heredia, Alanís-Guzmán & Arévalo-Niño, 2017) and improves thermal stability and flexibility (Guadarrama-Lezama *et al.*, 2018). Moreover, the combination of *Opuntia* mucilage and chitosan as a bioactive material improves the structure, stability and homogeneity of the film, as observed by SEM analysis and fundamental parameters (Maqbool, Ali, Alderson, Zahid & Siddiqui, 2011). The physicochemical and rheological characteristics of the CS-OM film in the present study are in accordance with those previously reported, where there is no standard trend in these parameters and they depend on the composition and proportions in each particular case (Razavi, Mohammad Amini & Zahedi, 2015; Saberi & Golding, 2018). However, maintaining the stability and the physicochemical characteristics of the film is important

prospective work, since these can modify the initial sensory characteristics of the product and affect the perception of potential buyers

The use of chitosan as an antifungal element is a widely explored field. However, the underlying mechanisms have not yet been fully elucidated. We propose a global mechanism that includes electrostatic interactions with the plasma membrane (the most accepted model) and a series of stress conditioning elements (such as the formation of reactive oxygen species) and a complex metabolic imbalance (Olicón-Hernández *et al.*, 2015; Olicón-Hernández, Uribe-Álvarez *et al.*, 2017). With respect to the effect of chitosan on *R. stolonifer*, it was observed that the polymer modified the germination of spores, affected fungal morphology, increased glucose consumption, inhibited H<sup>+</sup>-ATPase activity in the plasma membrane and caused a high potassium efflux (Dos Santos *et al.*, 2012; García-Rincón *et al.*, 2010; Guerra-Sánchez, Vega-Pérez, Velázquez-del Valle & Hernández-Lauzardo, 2009; Hernández-Lauzardo *et al.*, 2008; Hernández-Lauzardo *et al.*, 2011). An increase in the shelf-life of different fruits using edible CS-OM films has been observed in recent years, which is in agreement with the results of the present study (Ávila-Sosa *et al.*, 2012; Treviño-Garza *et al.*, 2017). However, in most of the cases, the films were mixed with essential oils or adjuvants to increase the antifungal effect (Yuan, Chen & Li, 2016) and few recent studies evaluate the effectiveness of these films under *in situ* or *in vitro* conditions.

As the results indicate, the presence of the polymer mixed with the *Opuntia* mucilage inhibited the proliferation of the fungus under *in vitro* and *in situ* conditions for the protection of tomatoes. These results are consistent with those reported by Martínez-Camacho *et al.*, who determined similar antifungal index values against *Aspergillus niger*. Nevertheless, *in situ* protection was not tested by them (Martínez-Camacho *et al.*, 2010). Recently, Robledo *et al.*, 2018, described thymol nanoemulsions incorporated into quinoa protein/chitosan edible films for the protection of cherry tomatoes, which significantly reduced fungal infection by *Botrytis cinerea* following incubation at 5 °C for seven days (Robledo *et al.*, 2018). In summary, the results obtained in the present study describe an attractive potential alternative that can be used under real conditions for the protection of fruits against infection with phytopathogenic fungi.

## CONCLUSION

An edible film was designed based on chitosan and *Opuntia* mucilage that presented a uniform and homogenous surface/structure. The best integrity was observed in the film containing the highest proportions of glycerol and chitosan, which affected its stability and antifungal effect. The CS-OM film significantly reduced the growth of *R. stolonifer* under

*in vitro* and *in situ* conditions for the protection of tomatoes and increased their shelf-life, making it a potential product for possible massive application in agriculture.

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## CONFLICT OF INTEREST STATEMENT

The authors declare that they have no conflict of interest.

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